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Cerambycidae and Disteniidae (Coleoptera) from Ecuador: new genera, new species, new records, and corrections in species-group names

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new genus;
new species;
South America.

Abstract. – Two new genera and three new species of Cerambycinae are described from Ecuador: *Kerasellum afasciatum*, **gen**. **nov**., **sp**. **nov**. (Tillomorphini); *Afti citrinum*, **gen**. **nov**., **sp**. **nov**. (Trachyderini); and *Compsibidion zonatum* **sp**. **nov**. (Tropidini), this latter is included in a previous key. Two species-group names in *Compsibidion* Thomson, 1864 are corrected and two new geographical records are reported. The current inclusion of *Eudereces elachys* Martins & Galileo, 2013 in *Tillomorphites* Vitali, 2011, is questioned in the remarks on *Kerasellum* **gen**. **nov**.

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Introduction

In recent years, the first author has been collecting regularly in different parts of Ecuador, which has revealed a considerable number of new species and new records. In some cases, it has been necessary to describe new genera to accommodate the new species. Currently, Ecuador has 1082 species and subspecies reported in its territory (Bezark 2024; Monné 2024a, b, c; Roguet 2025; Tavakilian & Chevillotte 2024).

American Tillomorphini (Cerambycidae, Cerambycinae) contains 15 genera and over 100 species distributed from Canada to southern South America, including the Caribbean (Bezark 2024; Monné 2024a; Monné & Nearns 2024; Tavakilian & Chevillotte 2024). Here we described one new genus and one new species from Ecuador.

American Trachyderini (Trachyderina) (Cerambycidae, Cerambycinae) has 132 genera and over 440 species distributed from Canada to southern South America, including the Caribbean (Bezark 2024; Monné 2024a; Monné & Nearns 2024; Tavakilian & Chevillotte 2024). Here we described one new genus and one new species from Ecuador.

Compsibidion Thomson, 1864 is a large genus of Tropidini (Neoibidionina) (Cerambycidae, Cerambycinae) with 86 species distributed from Mexico to southern South America, including the Caribbean, and seven species known from Ecuador (Bezark 2024; Monné 2024a; Tavakilian & Chevillotte 2024). Here we described a new species from Ecuador. Additionally, two species-group names in the genus are corrected.

Paracompsa Martins, 2009 (Cerambycidae, Tropidini, Compsina) includes two species occurring only in South America (Bezark 2024; Monné 2024a; Tavakilian & Chevillotte 2024). Here a new country record is reported.

Arietocometes Santos-Silva & Tavakilian, 2009 is a small genus of Disteniidae with nine species distributed from Mexico to northern South America (Bezark 2024; Monné 2024b; Tavakilian & Chevillotte 2024). Here a new province record in Ecuador is reported.

Materials and Methods

Photographs were taken with a Canon EOS Rebel T3i DSLR camera or Canon EOS R8 mirrorless camera, Canon MP-E 65mm f/2.8 1-5X macro lens; specimens were illuminated with Dome of light (RK Science Factory, Brazil). The specimens were examined with a stereomicroscope at 30x magnification. Measurements were taken in "mm."

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All specimens of the new species and those used to establish the new geographical records were collected by the first author and originally belonged to his private collection.
References on known species are restricted to the original descriptions; for full references see Monné (2024a, b) and Tavakilian & Chevillotte (2024).
The collection acronyms used in the text are as follows:

– **JVCO**: Josef Vlasak Collection, Schwenksville, Pennsylvania, U.S.A.
– **MZSP**: Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil.

Results

CERAMBYCIDAE Latreille, 1802

CERAMBYCINAE Latreille, 1802

TILLOMORPHINI Lacordaire, 1868

***Kerasellum* gen. nov.**

(Fig. 1-8)

ZooBank: https://zoobank.org/36C38EBA-C0F5-4299-9B16-B1A63639B71C

Type species. – *Kerasellum afasciatum* **sp. nov.**, here designated.

Etymology. – The name *Kerasellum* comes from Greek "κέρας" ("keras," meaning horn of an animal) with diminutive Latin suffix "ellum." Alluding to the centrobasal crest of the elytra, which resembles a small horn. Neuter gender.

Description

Female. – Small-sized body, somewhat ant-shaped. Body, antennae, and legs with long, erect, light-color setae. Head not retractile; frons distinctly transverse; maxillary palpomere IV and labial palpomere III securiform; antennal tubercles slightly elevated, well separated; genae about half maximum length of lower eye lobe in frontal view; antennae 11-segmented, slightly surpassing elytral apex; basal flagellomeres not tumid and without apical spine; distal antennomeres not strongly widened toward their apices; antennomere III shorter than scape, slightly shorter than V; upper eye lobes absent; lower eye lobes with small projection upward posterodorsally. Prothorax strongly narrowed posteriorly; pronotum densely, longitudinally vermiculate; prosternal process strongly narrowed centrally, moderately triangularly widened apically; procoxal cavities slightly open posteriorly. Elytra, in lateral view, depressed about middle, strongly elevated and convex on posterior half; apex rounded, unarmed; centrobasal crest elevated, narrow, C-shaped; without transverse light integumental band. Femora pedunculate-clavate; metatarsomere I slightly longer than II-III together.

Remarks. – Among the genera of Tillomorphini without distinct upper eye lobes (*Licracantha* Lingafelter, 2011; *Pentanodes* Schaeffer, 1904; and *Gourbeyrella* Lane, 1959), *Kerasellum* **gen. nov**. is similar to *Pentanodes*, but differs as follows: elytra without eburneous or yellowish transverse band; centrobasal crest of elytra C-shaped; and apex of prosternal process slightly widened, making procoxal cavities slightly open. In *Pentanodes*, the elytra have eburneous or yellowish transverse band (except in *P. albofasciatus* Fisher, 1932, provisionally kept in the genus by Heffern *et al.* (2020)), the centrobasal crest of elytra is longitudinal, from distinct to practically absent, and the apex of the prosternal process is strongly widened, making the procoxal cavities distinctly closed posteriorly. From *Gourbeyrella*, the new genus differs by the prothorax not subcylindrical (subcylindrical in *Gourbeyrella*), and elytra with C-shaped centrobasal crest (centrobasal crest absent in *Gourbeyrella*); and from *Licracantha* it differs by the antennomeres III-V without apical spine (present in *Licracantha*), elytra without eburneous or yellowish transverse band (present in *Licracantha*), and the centrobasal of elytra C-shaped (longitudinal and slightly tumid in *Licracantha*).
Vitali (2011) transferred *Eudereces elachys* Martins & Galileo, 2013) to *Tillomorphites* Vitali, 2011. However, this species has no upper eye lobes (Martins & Galileo 2013; one paratype male examined by us). The illustrations of the type species of the genus (Vitali 2011, 2017), *T. robustus* Vitali, 2011, and of *T. otiliae* Vitali, 2017 (see original description) clearly show that both species have distinct upper eye lobes. Therefore, we think that the transference of *Euderces elachys* to *Tillomorphites* was a mistake. A more detailed study will be necessary to correctly allocate *Euderces elachys*, which differs from *Kerasellum* by the absence of spine on the basal flagellomeres (present in *Tillomorphites elachys*).

On the tribal allocation of *Kerasellum*. – According to Martins (1975) (translated): "The systematic of the subfamily Cerambycidae at tribal level is almost incomprehensible and, in the groups related to Tillomorphini (*sensu* Lacordaire, 1869), seems to me to be one of the most chaotic, needing urgently a broad-spectrum review. One of the main confusions arises from the genus *Tillomorpha*, type of Tillomorphini; the type species of *Tillomorpha*, *T. lineoligera* Blanchard (by monotypy and later designation by Thomson 1860:229) [*sic*] deviates greatly from the tribal concept defined by Lacordaire (1869:88); in this species, the procoxal cavities are widely open behind and the mesocoxal cavities are visibly open laterally. Now, Lacordaire himself states that the Tillomorphini are a dismemberment of the Clytini, separable by the closed mesocoxal cavities, but, "il est, très-difficile de trouver une ligne de démarcation entre eux et ceux des groupes suivants (Cleomenini, Rhopalophorini, etc.) qui présentent le même caractère" [it is very difficult to find a line of demarcation between them and those of the following groups [Cleomenini, Rhopalophorini, etc.] which present the same feature] (p. 88)."
More recently, Heffern *et al.* (2020), reported: "The characteristics mentioned by diverse authors to separate Tillomorphini from Anaglyptini (and also Neocorini) appear not to be useful (see, for example, Lingafelter 2011). Thus, future studies to separate these tribes should be done."
Whether the new genus belongs to Tillomorphini or Anaglyptini (or even another tribe) remains opened. However, based on the general appearance much more similar to genera currently included in Tillomorphini, we prefer to include the new genus in this tribe.

***Kerasellum afasciatum* sp. nov.**

(Fig. 1-8)

ZooBank: https://zoobank.org/AA5A7415-501D-4F39-AEDD-ADB403722A78

Holotype, ♀, ECUADOR, *Napo*: Pacto Sumaco, 0°40'10"S 77°35'42"W, 23 Nov 2022, J. Vlasak leg. (MZSP, formerly JVCO).

Paratype, **1** ♀, same data as holotype (JVCO).

Description of the holotype (Fig. 1-7)

Coloration. – Integument mostly blackish; ventral mouthparts dark brown on mentum and basal area of labium, reddish brown on remaining segments, except extreme apex of maxillary palpomeres I-III and labial palpomeres I-II, and apical 2/3 of maxillary palpomere IV and labial palpomere III yellowish-brown; anteclypeus and labrum partially dark brown; mandible partially dark reddish brown between basal 2/3 and apical fourth; scape and pedicel dark brown except dark reddish-brown apex of dorsal surface; antennomere III reddish brown except dark-brown inner dorsal apex; antennomeres IV-V reddish brown except apical quarter mostly dark brown; antennomeres VI-VIII reddish brown except apical quarter; antennomeres IX-XI reddish brown basally, dark brown on remaining surface. Coxae mostly dark brown; trochanters mostly reddish brown; femora reddish brown, except dark-brown dorsal surface of femoral club; tibiae reddish brown basally and apically, dark brown on remaining surface; tarsomeres mostly reddish brown with irregular brown areas.

Head. – Frons densely rugose with sparse, coarse punctures interspersed; frontoclypeal sulcus deep, arched; with very sparse, short, erect white setae, and a few long, erect, thick yellowish-brown setae, especially laterally. Vertex and area behind antennal tubercles with sculpturing as on frons; with a few short, erect whitish setae and a few long, erect, thick yellowish-brown setae laterally between antennal tubercles, glabrous on remaining surface. Area

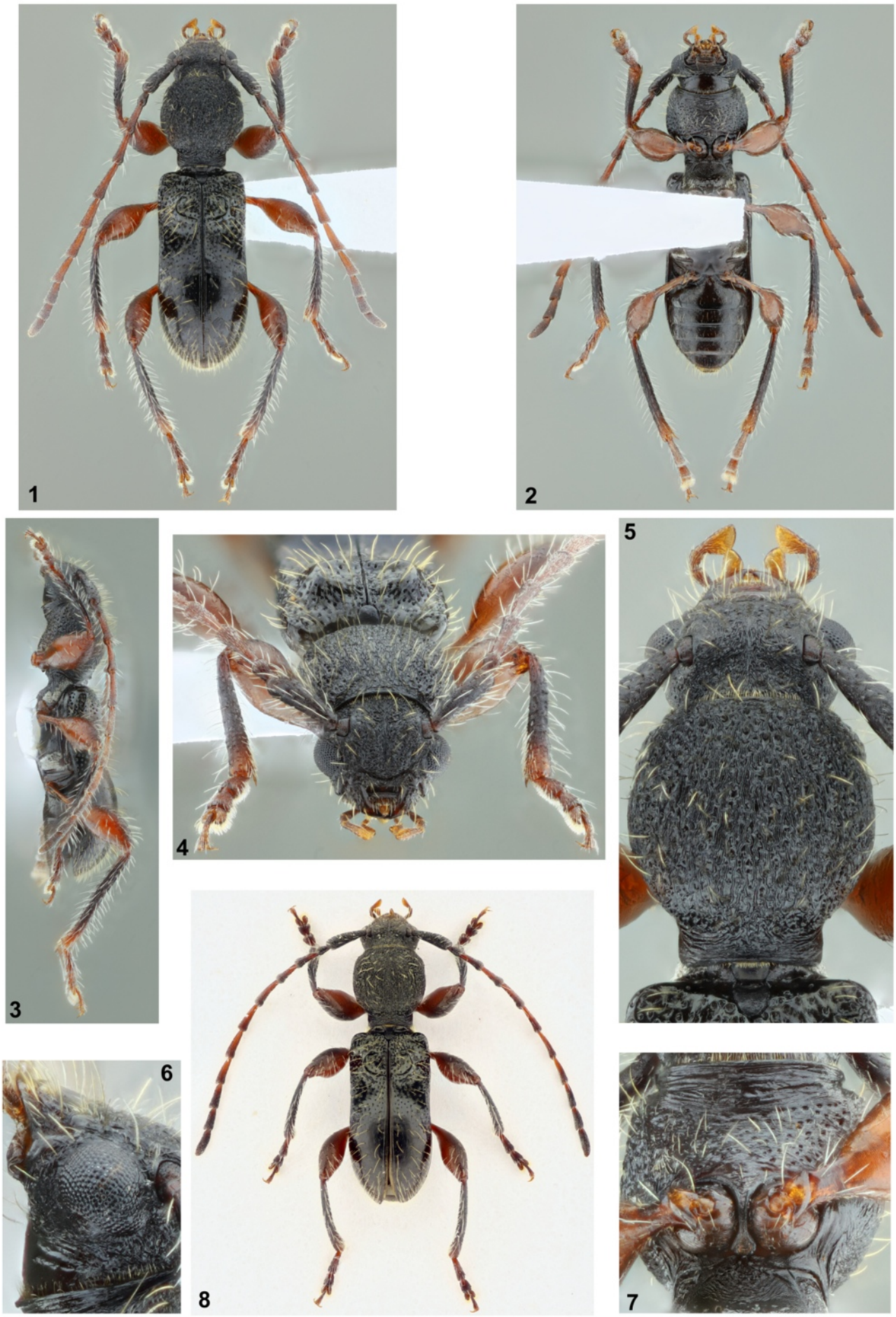

Fig. **1**-**8**. *Kerasellum afasciatum* **gen**. **nov**., **sp**. **nov**. **1**-**7**. Holotype ♀. **1**. Dorsal habitus. **2**. Ventral habitus. **3**. Lateral habitus. **4**. Head, frontal view. **5**. Head and pronotum. **6**. Eye. **7**. Procoxal cavities. **8**. Paratype ♂, dorsal habitus.

behind lower eye lobes slightly rugose close to eye, subsmooth close to prothorax; glabrous. Genae slightly rugose except smooth apex; with a few short, erect whitish setae toward ventral surface and long, erect, thick yellowish-brown setae, smooth area glabrous. Wide central area of postclypeus finely rugose; with short, bristly, sparse whitish setae and one long, erect, thick yellowish-brown seta on each side. Sides of postclypeus partially smooth, glabrous. Labrum coplanar with anteclypeus, finely, transversely striate on posterior third, inclined, subsmooth on remaining surface; glabrous on posterior third, with sparse, short, bristly whitish setae on central third, with long, erect yellowish setae interspersed, abundant, short light yellowish-brown setae on center of anterior third, glabrous on sides of anterior third. Posterior 2/3 of outer surface of mandibles coarsely rugose-punctate; with sparse, short whitish setae and long, erect, thick yellowish setae interspersed; apical third smooth, glabrous. Antennal tubercles with sculpturing as on frons except subsmooth apex; with sparse whitish setae. Posterior 2/3 of gulamentum smooth, glabrous; anterior third transversely striate-punctate; with a few long, erect light yellowish-brown setae. In frontal view, distance between lower eye lobes 0.61 times distance between outer margins of eyes. Antennae 1.7 times elytral length, surpassing elytral apex on basal quarter of antennomere XI. Scape very finely, irregularly striate, except smooth dorsal apex; sparsely, moderately coarsely punctate, except on dorsal apex; dorsal and lateral surfaces with a few short, decumbent white setae and long, erect, thick yellowish-white setae interspersed except glabrous dorsal apex; ventral surface with long, erect whitish setae on basal 2/3 and long, erect yellowish-white setae on apical third, yellowish-white setae longer than whitish setae. Pedicel minutely striate on basal 3/4, smooth on apical quarter; with a few short, decumbent whitish setae dorsally between middle and apical quarter, and long, erect whitish setae on apical half, except on smooth apical area. Antennomeres III-X gradually widened toward apex, more distinctly from IV to VIII; XI subcylindrical with narrowed, rounded apex. Antennomeres III-VI with moderately abundant whitish pubescence not obscuring integument, except glabrous base, and long, erect whitish setae ventrally and on apex of dorsal and lateral surfaces; with a few short, erect light yellowish-brown setae interspersed dorsally, setae gradually more abundant toward VI. Antennomeres VII-XI with abundant whitish pubescence not obscuring integument, gradually denser toward XI, and short, erect yellowish setae interspersed throughout; VII-VIII with a few long whitish setae on apex of dorsal surface; VII with a few long erect whitish setae on apex of ventral surface.

Antennal formula (ratio) based on length of antennomere III:
– Scape = 1.50. – Pedicel = 0.45. – IV = 1.10. – V = 1.30. – VI = 1.05. – VII = 1.00. – VIII = 0.95. – IX = 0.90. – X = 0.70. – XI = 0.95.

Thorax. – Sides of prothorax rounded from anterolateral angles to posterior sixth, then parallel-sided toward posterolateral angles. Anterior 5/6 of pronotum densely, irregularly, longitudinally vermiculate, with coarse punctures interspersed; sides of posterior sixth transversely striate, less so toward posterolateral angles, central area rugose; anterior 5/6 with sparse, long, erect, thick yellowish-brown setae, sparser centrally; posterior sixth glabrous. Sides of prothorax with sculpture similar to that on wide central area of pronotum, but punctures more abundant, except subsmooth area close to anterior and posterior margins; with sparse, long, erect, thick light yellowish-brown setae on wide central area, especially anteriorly. Anterior 1/3 of prosternum transversally striate with sparse grayish-white pubescence centrally. Posterior 2/3 of prosternum moderately abundantly, coarsely punctate laterally, densely, finely, transversely striate-punctate centrally; sides with moderately sparse grayish-white pubescence and central area with sparse, long, erect, thick yellowish-white setae. Prosternal process with sides elevated; apex slightly rounded; finely rugose-punctate centrally, smooth laterally; glabrous; narrowest area 0.13 times procoxal width. Sides of anterior half of central area of mesoventrite densely, finely, obliquely striate, glabrous; remaining surface of anterior half finely rugose, glabrous; posterior half of central area sparsely, coarsely punctate, with a few, both short and long, erect yellowish setae; sides with abundant whitish pubescence obscuring integument, except glabrous apex. Mesoventral process with sparse, long, erect yellowish setae. Mesanepisterna finely rugose; with a few long, bristly yellowish-white setae close to central area of mesoventrite and dense whitish pubescence close to elytra. Mesepimera glabrous. Metanepisterna mostly glabrous on anterior 3/4 and dense whitish pubescence over most of posterior quarter. Metaventrite with dense whitish pubescence close to metacoxal cavities, and a few long, erect, thick yellowish setae on remaining surface. Scutellum minutely striate, glabrous, with rounded apex.

Elytra. – Abundantly, coarsely punctate on anterior quarter, punctures sparser, finer on remaining anterior half, and very sparse and fine on posterior half; with sparse, long, erect, thick yellowish-brown setae, setae slightly more abundant on anterior quarter, and abundant, long, erect yellowish setae on sides and apex of posterior quarter.

Legs. – Femora abundantly, minutely striate on dorsal and lateral surfaces of femoral club; sparsely, coarsely punctate throughout; with a few short, decumbent yellowish setae throughout, and sparse, long, erect, thick yellowish-white setae throughout. Tibiae abundantly, minutely striate, dorsal and lateral surfaces with moderately abundant, long, erect, thick yellowish-white setae; ventral surface with sparse yellowish-white setae, setae gradually longer toward area near apex, except apex with abundant, bristly yellowish-brown setae. Dorsal surface of tarsomeres with whitish pubescence not obscuring integument and long, erect setae of same color interspersed.

Abdomen. – Ventrites very sparsely and finely punctate; with a few, both short, decumbent whitish setae and more abundant, long, erect yellowish setae, and yellowish-brown on sides of ventrite 5; apex of ventrite 5 rounded.

Dimensions (mm) (holotype ♀ – paratype ♀)
– Total length, 5.10-5.40;
– Prothoracic length, 1.35-1.55;
– Anterior prothoracic width, 0.90-0.95;
– Posterior prothoracic width, 0.75-0.85;
– Maximum prothoracic width, 1.25-1.35;
– Humeral width, 1.30-1.45;
– Elytral length, 3.00-3.20.

Etymology. – The name "afasciatum," from Latin "fasciatus" (banded), with a prefix "a" (without) alludes to the lack of the eburneous transverse band, which is present in most species of Tillomorphini.

Remarks. – See remarks in the description of the genus.

TRACHYDERINI Dupont, 1836

TRACHYDERINA Dupont, 1836

***Afti* gen. nov.**

(Fig. 9-13)

ZooBank: https://zoobank.org/AEF4931E-334A-44C0-A385-67288B68883C

Type species. – *Afti citrinum* **sp. nov**., here designated.

Etymology. – Greek, "αφτί" (aftí), meaning "ear." Alluding to the shape of the lateral tubercles of the prothorax, the most conspicuous feature of the genus. Neuter gender.

Description

Female. – Moderately large-sized body, not flattened, somewhat slender. Head without dense pubescence and/or long erect setae; frons proportionally short; postclypeus large; frontoclypeal sulcus well marked; sensory area of apical palpomeres well marked; antennal tubercles moderately elevated, well separated; genae slightly shorter than maximum diameter of lower eye lobe; area behind upper eye lobes about as long as maximum diameter of one upper lobe; mandible shorter than width of postclypeus; eye lobes not separated; antennae 11-segmented, distinctly shorter than elytra, unicolorous; scape without dorsal depression, not flattened dorsally; antennomeres III-XI not flabellate, without dorsal sulcus, without fringe of setae ventrally and tuft of setae; antennomeres gradually shorter from V to X; antennomere III as long as scape, longer than IV; antennomere IV slightly shorter than V; apex of antennomeres III-X without apical spiniform projection. Prothorax transverse; anterior margin distinctly narrower than posterior one; lateral tubercles large, oblique truncate apically, located about middle; posterior area rounded projected. Pronotum opaque, densely, moderately finely, shallowly punctate except smooth central gibbosity; with distinctly, circular gibbosity on each side of anterior half and elongated, slightly elevated gibbosity centrally on posterior 2/3; without pubescence and erect setae. Prosternum transversely concave. Prosternal process slightly vertical apically. Mesoventral process distinctly wider than prosternal process,

Fig. **9**-**13**. *Afti citrinum* **gen**. **nov**., **sp**. **nov**., holotype ♀. **9**. Dorsal habitus. **9a**. Dorsal habitus showing the original color of the holotype. **10**. Ventral habitus. **11**. Lateral habitus. **12**. Head, frontal view. **13**. Detail of the lateral tubercle of prothorax.

Fig. **14**-**17**. *Athetesis prolixa* Bates, 1870, ♀. **14**. Dorsal habitus. **15**. Ventral habitus. **16**. Lateral habitus, anterior region of the body. **17**. Head, frontal view.

with lateroapical flaps; not tuberculate and not projected forward on anterior area; apical width slightly shorter than mesocoxa. Scutellum moderately small, about as long as half length of antennomere III, with rounded apex. Elytra about four times longer than prothorax; mostly parallel-sided; opaque, without eburneous callosities; abundantly, finely punctate; without distinct longitudinal dorsal and humeral carinae; apex unarmed, together rounded; without long, erect setae. Meso- and metafemora not pedunculate-clavate; apices without spines. Metatarsomere I longer than II-III together.

Remarks. – *Afti* **gen**. **nov**. is somewhat similar to *Athetesis* Bates, 1870 (Fig. 14-17), but differs as follows: body proportionally wider; lateral tubercles of prothorax obliquely truncate apically; pronotal and elytral punctures fine; prosternum transversely concave; elytra without distinct longitudinal carinae dorsally; and inner apex of meso- and metafemora rounded. In *Athetesis*, the body is proportionally narrower, lateral tubercles of the prothorax are subacute apically, pronotal and elytral punctures are coarse, prosternum not transversely concave, elytra have distinct longitudinal carinae dorsally, and the inner apex of the meso- and metafemora has short spiniform projection. The new genus is also similar to *Amphionthe* Bates, 1879 but differs especially by the antennae distinctly shorter than elytra in female (longer in females of *Amphionthe*), and by the shape of the lateral tubercles of the prothorax (conical in *Amphionthe*); from females of *Crioprosopus* Audinet-Serville, 1834 by the antennae distinctly shorter than the elytral length (about as long in females of *Crioprosopus*), shape of the lateral tubercles of the prothorax (conical in females of *Crioprosopus*), and scutellum not triangular (triangular in *Crioprosopus*); from *Gambria* Chevrolat, 1863 by the antennae distinctly shorter than the elytral length (about as long in females of *Gambria*) and the shape of the lateral tubercles of the prothorax (conical in females of *Gambria*); from females of *Megapurpuricenus* Eya, 2015 especially by the pronotal and elytral sculpturing much less distinct (densely punctate in *Megapurpuricenus*) and the shape of the lateral tubercles of the prothorax (conical in *Megapurpuricenus*); from *Neoxoplus* Eya, 2019 by the shape of the lateral tubercles of the prothorax (not truncate in *Neoxoplus*), procoxal process distinctly narrower than the procoxal cavity (about as wide as 2/3 of the procoxal width in *Neoxoplus*), and the apex of the scutellum rounded (acute in *Neoxoplus*); from *Parevander* Aurivillius, 1912 by the shape of the lateral tubercles of the prothorax (subacute in *Parevander*) and by the elytra without distinct longitudinal carinae dorsally (present in

Parevander); from *Paroxoplus* Chemsak, 1959 by the antennae in female much shorter than the elytra (about as long in females of *Paroxoplus*) and by the shape of the lateral tubercles of prothorax (subacute in *Paroxoplus*); and from *Phaedinus* Dupont, 1834, a genus with specimens sharing some features but strongly different in others, by the shape of the lateral tubercles of the prothorax (subacute in *Phaedinus*) and scutellum short and with rounded apex (long and with acute apex in *Phaedinus*). Additionally, it differs from some species of *Eriphus* Audinet-Serville, 1834 especially by the shape of the lateral tubercles of the prothorax (subacute in some species of *Eriphus*).

Afti citrinum **sp**. **nov**.

(Fig. 9-13)

ZooBank: https://zoobank.org/EE8747D7-8314-40A6-B748-BDDE75BC8456

Holotype, ♂, ECUADOR, *Pichincha*: Milpe Bird Sanctuary, 1100 m, 7 Jul 2023, J. Vlasak leg. (MZSP, formerly JVCO).

Description of the holotype (Fig. 9-13)

Coloration. – Head capsule black except ventral surface light brown with sides of posterior half orangish; ventral mouthparts mostly dark brown, except light brown base of mentum and yellowish apex of palpomeres; anteclypeus partially dark brown; mandibles black; antennae black, more dark brown toward apical segments. Prothorax light orange, except dark-brown area close to procoxal cavities and wide central area of prosternal process. Ventral surface of meso- and metathorax dark brown. Scutellum and elytra orange, except brown macula near elytral apex. Coxae dark brown; femora, tibiae, and tarsi black. Ventral surface of abdomen dark brown.

Head. – Frons densely, finely punctate anterocentrally, punctures slightly sparser and coarser laterally and posteriorly; punctures with minute dark-brown setae. Median groove well marked from clypeus to area between upper eye lobes. Area between antennal tubercles excavated centrally; densely, moderately coarsely punctate on sides of excavated area and close to antennal tubercles, subsmooth on center of excavated area; a few punctures with minute dark-brown setae. Area between antennal tubercles and center of area between upper eye lobes abundantly, coarsely punctate, except smooth central

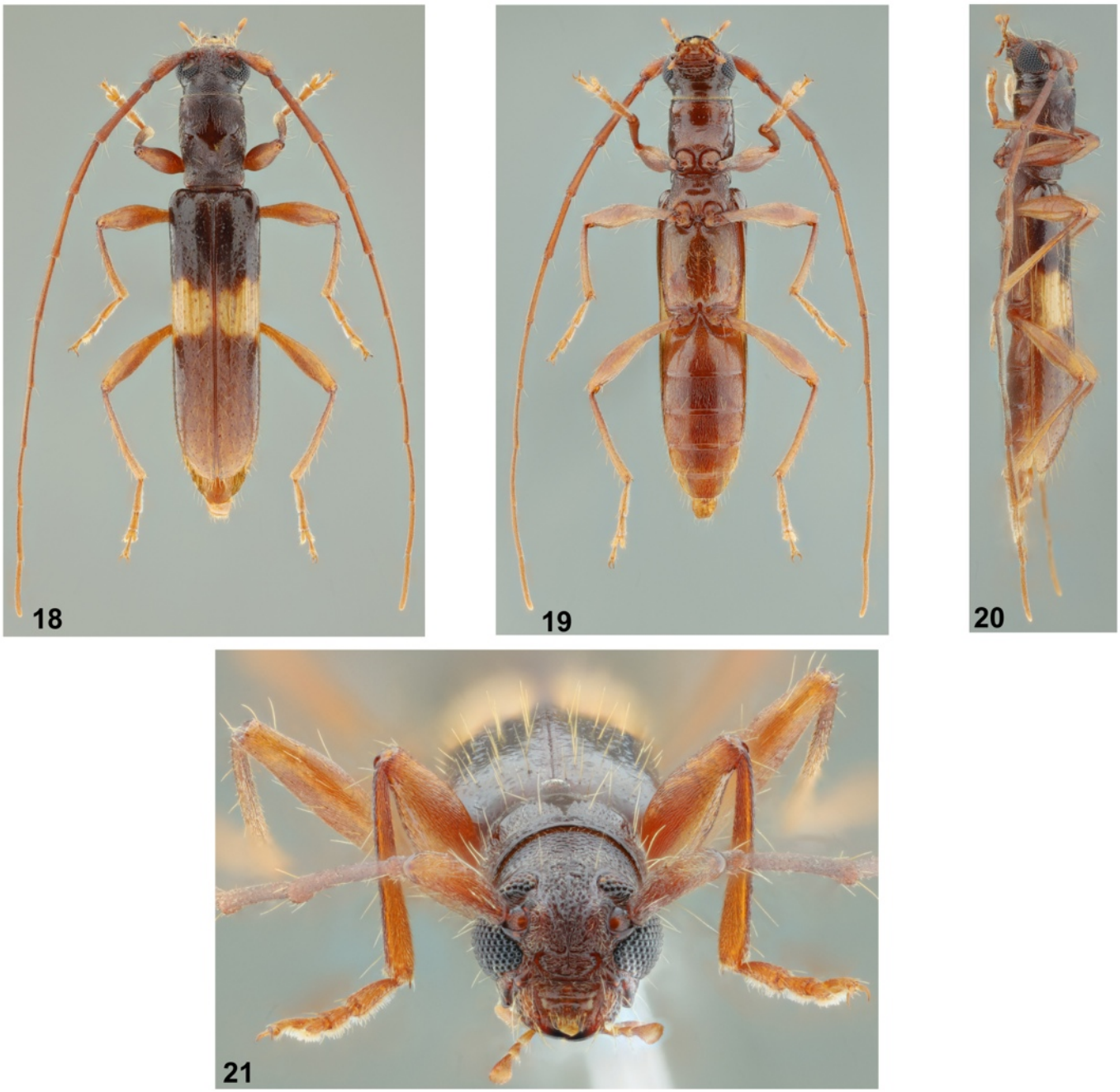

Fig. **18**-**21**. *Compsibidion zonatum* **sp**. **nov**., holotype male. **18**. Dorsal habitus. **19**. Ventral habitus. **20**. Lateral habitus. **21**. Head, frontal view.

area. Remaining surface of vertex densely, moderately coarsely punctate. Area behind upper eye lobes abundantly, coarsely punctate, except smooth inferior area close to eye; with one long, erect dark yellowish-brown seta close to superior area of eye. Area behind lower eye lobes densely, moderately finely punctate close to eye, punctures slightly sparser and coarser close to prothorax, and shallower and sparser centrally. Genae densely, moderately coarsely punctate toward ventral surface, slightly sparser toward frons and clypeus. Tentorial pits well marked. Postclypeus densely, finely punctate close to frons, subsmooth on remaining posterior half, anterior half densely, finely punctate close to subsmooth area, and moderately sparsely, finely punctate close to anteclypeus; punctures on anterior half with minute yellowish seta. Sides of postclypeus smooth, glabrous. Labrum coplanar with anteclypeus posteriorly, inclined on remaining surface, more abruptly on anterocentral region; moderately sparsely, finely, shallowly punctate close to anteclypeus, abundantly punctate on remaining wide central area, subsmooth on remaining surface; punctures on posterior third with minute brownish seta, punctures on middle of wide central area with minute brownish seta except a few punctures with long, erect yellowish-brown seta, and punctures on sides of wide central area with abundant, long, erect, thick dark-brown seta; anterior region glabrous except apex of central region with fringe of yellowish-brown setae. Outer surface of mandibles abundantly, coarsely punctate on basal half, smooth on apical half; punctures with minute yellowish-brown seta, except punctures close to smooth area with long, erect, thick yellowish-brown seta. Gulamentum smooth, glabrous on posterior half, except transversely striate sides; anterior half abundantly, finely punctate laterally, striate-punctate centrally, except subsmooth intermaxillary process, with long, erect yellowish-brown setae laterally, mostly glabrous centrally. Antennal tubercles abundantly, moderately coarsely punctate basally, smooth apically. Distance between upper eye lobes 0.44 times distance between outer margins of eyes; in frontal view, distance between lower eye lobes 0.65 times distance between outer margins of eyes. Antennae 0.7 times elytral length, slightly surpassing anterior 2/5 of elytra. Scape abundantly, finely punctate, except smooth dorsal apex; punctures with minute brown seta, except on apical region of inner and ventral surfaces with short, erect dark-brown seta. Pedicel moderately abundantly, finely punctate, except smooth basal and apical surfaces; punctures with minute dark-brown seta, except a few punctures on ventral surface with short, erect dark-brown seta. Antennomeres III-XI densely, finely punctate, punctures denser and minute from VI, except smooth dorsal apex of III-VI; with abundant dark-brown pubescence not obscuring integument, pubescence denser from VI; ventral surface of III-VI with sparse, short, erect dark-brown setae, setae more abundant apically; apex of dorsal surface of III-X with moderately abundant, short, erect dark-brown setae; apex of ventral surface of VII-X with sparse, short, erect dark-brown setae; XI with sparse, short, erect dark-brown setae.

Antennal formula (ratio) based on length of antennomere III:
– Scape = 1.00. – Pedicel = 0.20. – IV = 0.83. – V = 0.87. – VI = 0.75. – VII = 0.77. – VIII = 0.63. – IX = 0.63. – X = 0.50. – XI = 0.67.

Thorax. – Pronotum glabrous, except a few short, erect pale-yellow setae posterolaterally. Sides of prothorax abundantly, shallowly, moderately coarsely punctate, except smooth area close to anterior and posterior margins and area close to procoxal cavities; glabrous. Prosternum moderately abundantly, finely, shallowly punctate on posterior 2/3, punctures partially asperate, with a few short, erect brown setae posterocentrally and close to procoxal cavities; anterior third abundantly, finely, shallowly punctate laterally, slightly transversely striate on wide central area, except smooth, slightly depressed area close to anterior margin, glabrous except one moderately long, erect brown seta on each side. Prosternal process slightly depressed centrally; abundantly, finely punctate; with sparse yellowish-white setae; narrowest area 0.32 times procoxal width. Mesoventrite densely, moderately finely punctate, except subsmooth apices; with abundant, bristly brown pubescence not obscuring integument, pubescence indistinct depending on viewing angle, with a few short, erect brown setae interspersed, except subsmooth apices with sparse, decumbent yellowish-white setae. Mesanepisterna and mesepimera densely asperate-punctate; pubescence and erect setae as on central area of mesoventrite, except fringe of dull yellowish-brown setae on posterior margin of mesepimera. Mesoventral process densely, moderately finely punctate, except subsmooth apical tabs; with abundant brown pubescence not obscuring integument, pubescence indistinct depending on viewing angle, and short, erect brown setae interspersed, except subglabrous apical tabs and posterocentral area with moderately abundant, short yellowish-brown setae directed backward; narrowest area 0.68 times mesocoxal width. Metanepisterna with abundant brown pubescence not obscuring integument, pubescence indistinct depending on viewing angle, and short, erect brown setae interspersed. Metaventrite densely, finely punctate laterally, abundantly, moderately coarsely centrally, except subsmooth area close to metacoxal cavities; with abundant brown pubescence laterally not obscuring integument, pubescence sparser centrally, absent close to metathoracic discrimen, and short, erect, abundant yellowish-brown setae interspersed, pubescence and erect setae indistinct depending on viewing angle. Scutellum abundantly, minutely punctate except smooth sides; with moderately abundant, minute brown setae centrally, absent laterally.

Elytra. – Glabrous, except dense fringe of yellowish setae on epipleural margin and short, erect yellowish-brown setae on posterior sixth, setae more abundant close to apex.

Legs. – Femora abundantly, moderately coarsely asperate-punctate, punctures slightly sparser apically; a few punctures on profemora with short dark-brown seta; meso- and metafemoral punctures on dorsal and lateral surfaces with short, decumbent dark-brown seta except a few punctures with short, erect dark-brown seta; meso- and metafemoral punctures on ventral surface with short, erect dark-brown seta. Tibiae densely, finely asperate-punctate, punctures slightly sparser on protibiae and absent close to apex of sides of meso- and metatibiae; with abundant, bristly, short dark-brown setae not obscuring integument, setae denser ventrally, especially apically, except glabrous smooth areas on meso- and metatibiae. Tarsomeres I-III with abundant, decumbent dark-brown setae dorsally; IV-V with sparse dark-brown setae.

Abdomen. – Ventrites with abundant, bristly yellowish-brown pubescence, indistinct depending on viewing angle, with slightly longer, erect setae interspersed, except glabrous central apex of ventrites 1, 2, and 4, and partially glabrous central apex of ventrite 3; apex of ventrite 5 truncate.

Dimensions (mm)

– Total length, 20.35;
– Prothoracic length, 3.30;
– Anterior prothoracic width, 2.60;
– Posterior prothoracic width, 4.00;
– Maximum prothoracic width, 5.15;
– Humeral width, 5.40;
– Elytral length, 15.50.

Etymology. – The name "citrinum" (Latin, meaning "lemon yellow") alludes to the color of the beetle.

Remarks. – The holotype was found in a large living branch that broke off and fell because of the galleries the beetled excavated, suggesting that this species develops in living wood, which is not uncommon among Trachyderini. Originally, the beetle was distinctly lemon yellow (Fig. 9a), which is why we named the species "citrinum," but the color faded over time.

TROPIDINI Martins & Galileo, 2007

NEOIBIDIONINA Monné, 2012

Compsibidion Thomson, 1864

***Compsibidion zonatum* sp. nov.**
(Fig. 18-25, 30-36)

ZooBank: https://zoobank.org/991D771C-DF85-4C43-A3BB-8AB10464BAD2

Holotype, ♂, ECUADOR, *Loja*: Rumi Wilco Ecolodge, 1600 m, Vilcabamba, 11 Aug 203, J. Vlasak leg. (MZSP, formerly JVCO).

Paratypes, **2** ♀, same data as holotype (JVCO).

Description of the holotype (Fig. 18-21, 36)

Coloration. – Head capsule reddish brown ventrally, on sides of frons, and frontoclypeal sulcus, dark brown on most central area of frons, dark brown on vertex and behind eyes, reddish brown on antennal tubercles except apices partially dark brown; ventral mouthparts light reddish brown, except orangish

Fig. **22**-**25**. *Compsibidion zonatum* **sp**. **nov**., paratypes ♀. **22**-**23**. Specimen 2. **22**. Dorsal habitus. **23**. Ventral habitus. **24**-**25**. Specimen 1. **24**. Dorsal habitus. **25**. Ventral habitus.

apex of palpomeres; clypeus and labrum mostly reddish brown; antennae light reddish brown, more orangish brown depending on light intensity. Prothorax dark brown dorsally and laterally, reddish brown ventrally. Ventral surface of meso- and metathorax reddish brown, except dark-brown area close coxal cavities and darker on lateral and posterior margins of metaventrite. Scutellum dark brown. Elytra dark brown on anterior third, except dark reddish-brown circum-scutellar area and light reddish-brown epipleural margin, yellowish brown between anterior third and middle, this area orangish close to suture, and reddish brown, gradually yellowish brown toward apex on posterior half. Legs mostly orangish brown, more light reddish brown depending on light intensity, with irregular, slightly darker areas. Abdominal ventrites light reddish brown. See remarks.

Head. – Frons densely, coarsely punctate, punctures partially anastomosed; frontoclypeal sulcus deep, semi-circular; frontal plate well marked, straight on posterior margin, semi-circular on remaining margins, abundantly, coarsely punctate, somewhat striate-punctate posteriorly; with irregular Y-shaped shallow sulcus from frontal plate to area between antennal tubercles; with moderately abundant, short yellowish setae not obscuring integument laterally and a few short yellowish setae on remaining surface. Area between antennal tubercles and upper eye lobes abundantly, coarsely punctate, except smooth, slightly elevated longitudinal carina centrally; with sparse yellowish-white pubescence. Remaining surface of vertex abundantly, coarsely asperate-punctate on wide triangular central area, slightly rugose-punctate laterally toward eyes; with moderately sparse yellowish-white pubescence on its anterior region, glabrous close to prothorax; with one long, erect yellowish-brown seta close to eyes. Area behind upper eye lobes slightly rugose close to eye, except smooth area close to lower eye lobe, slightly asperate-punctate close to prothorax; with sparse yellowish-white pubescence superiorly near eye, glabrous on remaining surface. Area behind lower eye lobes sparsely, coarsely punctate, slightly rugose on some regions; with long, erect yellowish-brown setae near eyes, slightly more

abundant toward ventral surface. Genae abundantly, coarsely punctate, except smooth apex; with moderately sparse yellowish pubescence, except glabrous apex; with a few long, erect yellowish-white setae. Antennal tubercles with apex triangular, slightly elevated; with moderately abundant yellowish pubescence not obscuring integument. Area between antennal base and eyes with dense yellowish pubescence. Wide central area of postclypeus with sparse, short, bristly yellowish setae, absent on middle close to frons, and one long, erect yellowish-white seta on sides. Sides of postclypeus smooth, glabrous. Labrum smooth, glabrous close to anteclypeus; central area moderately abundantly and coarsely punctate, with moderately abundant, both short and long yellowish-brown setae; anterior area with abundant, short yellowish-brown setae directed forward, partially obscuring integument. Gulamentum smooth, glabrous on posterior half; anterior half moderately abundantly, coarsely punctate, with two transverse carinae, one about middle, another close to intermaxillary process, with sparse, short yellowish setae, and long, erect setae of same color interspersed. Basal 2/3 of outer surface of mandibles abundantly, coarsely punctate, with short, decumbent yellowish-brown setae not obscuring integument, and long, erect setae of same color interspersed; apical third smooth, glabrous. Upper eye lobes with three rows of ommatidia, except two ommatidia on inner apex. Distance between upper eye lobes 0.30 times distance between outer margins of eyes; in frontal view, distance between lower eye lobes 0.46 times distance between outer margins of eyes. Antennae 2.1 times elytral length, surpassing elytral apex at apical third of antennomere VIII. Scape with abundant yellowish-white pubescence not obscuring integument, except glabrous dorsal apex; with a few long, erect yellowish setae dorsally, laterally, and close to apex of ventral surface. Pedicel with abundant yellowish-white pubescence not obscuring integument, and a few long, erect yellowish setae throughout. Antennomeres III-XI, not carinate dorsally, not widened, with abundant yellowish-white pubescence not obscuring integument; III-V with a few long, erect yellowish setae ventrally and on apex of dorsal and lateral surfaces; VI-X with long, erect yellowish setae on apex of ventral surface, setae gradually shorter toward X; VI with a few long, erect yellowish setae on apex of dorsal surface.

Antennal formula (ratio) based on length of antennomere III:
– Scape = 0.64. – Pedicel = 0.19. – IV = 0.87. – V = 0.97. – VI = 0.97. – VII = 0.97. – VIII = 0.90. – IX = 0.84. – X = 0.77. – XI = 0.80.

Thorax. – Prothorax longer than wide, cylindrical, with sides slightly concave from after middle to posterior sixth. Pronotum with slightly elevated gibbosity on sides slightly before middle, elongated, slightly elevated gibbosity centrally, between anterior and posterior thirds, and another small, slightly elevated gibbosity on each side of posterior fifth; abundantly, minutely punctate, except smooth cross-shaped central area; with abundant yellowish pubescence not obscuring integument, except glabrous smooth area; with a few long, erect yellowish-brown setae. Sides of prothorax abundantly, minutely punctate except slightly transversely striate area close to anterior and posterior margins; with abundant yellowish pubescence not obscuring integument, except glabrous striate areas; with a few long, erect yellowish-brown setae interspersed on pubescent region. Prosternum moderately abundantly, minutely punctate on sides of posterior 3/4 and close to procoxal cavities, mostly smooth on remaining surface; with moderately abundant yellowish pubescence not obscuring integument on sides of posterior third and centrally close to prosternal process, and a few short, decumbent yellowish setae on remaining surface; with one long, erect yellowish-brown seta on each side of area close to prosternal process. Prosternal process with a few short, decumbent yellowish setae on anterior half, glabrous on remaining surface; narrowest area 0.08 times procoxal cavity. Mesoventrite with moderately sparse yellowish pubescence. Mesanepisterna, mesepimera, metanepisterna, and sides of metaventrite with abundant yellowish-white pubescence not obscuring integument; remaining surface of metaventrite with sparse yellowish-white pubescence, except glabrous area close to metathoracic discrimen, and a few long, erect yellowish setae interspersed. Mesoventral process distinctly widened on sides of apical region; posterior margin distinctly notched centrally; with abundant yellowish pubescence not obscuring integument, pubescence sparser apically, and a few long, erect yellowish setae interspersed; narrowest area 0.49 times mesocoxal cavity; widest area 0.75 times mesocoxal cavity. Scutellum with abundant yellowish pubescence not obscuring integument.

Elytra. – Sparsely, moderately sparsely punctate; not pubescent, with long, erect yellowish-brown seta in some punctures, slightly more abundant on posterior quarter; apex rounded toward sutural angle, without projections.

Legs. – Femora, tibiae, and dorsal surface of tarsi with abundant yellowish-white pubescence not obscuring integument, except dense, bristly yellowish pubescence on inner apical surface of protibiae, apical half of ventral surface of protibiae, and apical third of meso- and metatibiae; with long, erect yellowish setae, slightly more abundant on metatibiae; metatarsomere I slightly longer than II-III together.

Abdomen. – Ventrites somewhat abundantly, minutely punctate, except smooth apex of ventrites 1-4; with abundant yellowish pubescence not obscuring integument, except glabrous smooth areas, and a few long, erect yellowish-white setae interposed; apex of ventrite 5 truncate, slightly notched centrally.

Female (Fig. 22-25, 30-35)
– Similar to male, antennae shorter, 1.5-1.7 times elytral length, reaching elytral apex between antennomeres X and XI (proportions slightly varied between the two females);
– Abdominal ventrite 5 distinctly narrower.

Chromatic variations in the specimens examined. – Head capsule entirely brown; scape orangish brown; pedicel and flagellomeres brown; prothorax reddish brown, lighter ventrally; anterior third and posterior half of elytra dark brown, except yellowish-brown sides of posterior quarter and apex; yellowish-brown area on elytra surpassing middle; posterior third of metaventrite orangish brown; femora with large and irregular brownish areas.

Morphological variations in the specimens examined. – Anterior third of prosternal process transversely striate; narrowest area of prosternal process in females from 0.16 to 0.24 times procoxal width; sides of mesoventrite with abundant yellowish pubescence partially obscuring integument; elytra with very sparse, minute, decumbent yellowish-white setae.

Dimensions (mm) (holotype ♂ / paratypes ♀)
– Total length, 9.50/9.10-11.10;
– Prothoracic length, 1.70/1.80-2.25;
– Anterior prothoracic width, 1.20/1.25-1.50;
– Posterior prothoracic width, 1.15/1.15-1.25;
– Maximum prothoracic width, 1.35/1.35-1.75;
– Humeral width, 1.75/1.85-2.20;
– Elytral length, 5.70/5.80-7.10.

***Compsibidion zonatum* sp. nov. can be included in the alternative of couplet "4" in the key from Santos-Silva *et al.* (2017)**

4(2). Upper eye lobes with 5 rows of ommatidia on widest area; elytra at least partially with moderately abundant, decumbent pubescence. Bolivia ..... *C. amboroense* Santos-Silva, Galileo & Wappes, 2017
— Upper eye lobes with 3 rows of ommatidia on widest area; elytra with very sparse, decumbent pubescence ..... **4'**

4'(4). Inner apex of upper eye lobes with three ommatidia; central macula on elytra yellowish-white; posterior area of elytra dark brown. Bolivia ..... *C. achiraense* Santos-Silva, Galileo & Wappes, 2017
— Inner apex of upper eye lobes with two ommatidia; central macula on elytra yellowish brown; posterior area of elytra at least partially yellowish brown. Ecuador ..... *C. zonatum* **sp. nov.**

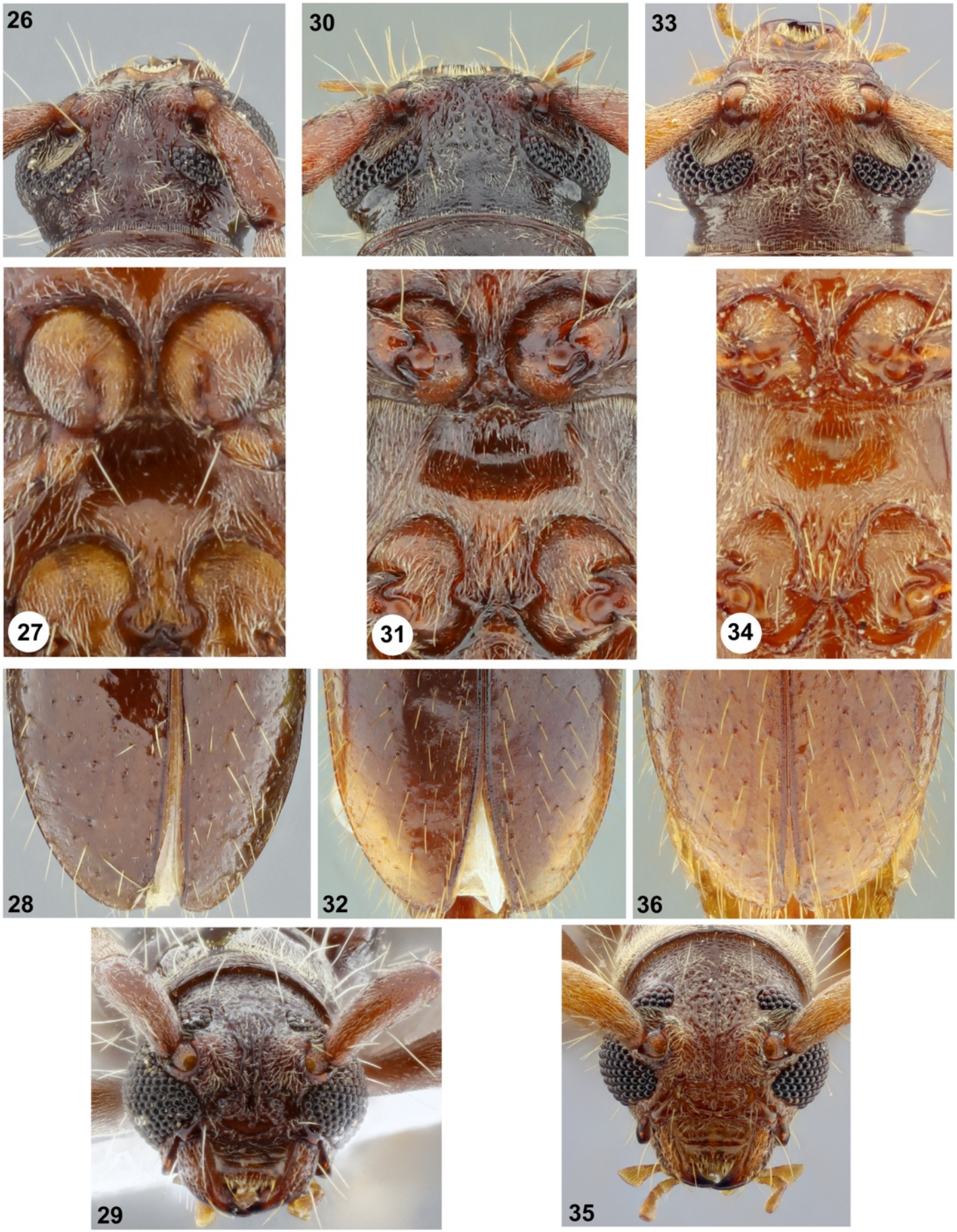

Fig. **26**-**36**. *Compsibidion* spp. **26**-**29**. *Compsibidion achiraense* Santos-Silva, Galileo & Wappes, 2017, holotype female. **26**. Head, dorsal view. **27**. Prosternal and mesoventral processes. **28**. Posterior area of elytra. **29**. Head, frontal view. **30**-**32**. *Compsibidion zonatum* **sp**. **nov**., paratype ♀, specimen 1. **30**. Head, dorsal view. **31**. Prosternal and mesoventral processes. **32**. Posterior area of elytra. **33**-**35**. *Compsibidion zonatum* **sp**. **nov**., paratype female, specimen 2. **33**. Head, dorsal view. **34**. Prosternal and mesoventral processes. **35**. Head, frontal view. **36**. *Compsibidion zonatum* **sp**. **nov**., holotype ♂, posterior area of elytra.

Etymology. – The name "zonatum" (Latin, meaning banded) refers to the three color zones (brownish – yellowish – brownish) on the elytra.

Remarks. – *Compsibidion zonatum* **sp. nov**. is similar to *C. achiraense* Santos-Silva, Galileo & Wappes, 2017 (Fig. 26-29; see also photographs on Bezark (2025) and in Santos-Silva *et al.* (2017)), but differs as follows: in females, distance between upper eye lobes wider, 0.36 times distance between outer margins of eyes (Fig. 30, 33); in females, distance between lower eye lobes 0.5 times distance between outer margins of eyes, making frons wider (Fig. 35); longitudinal carinae between antennal tubercles almost absent in both sexes (Fig. 30, 33, 35); in females, narrowest area of mesoventral process distinctly wider than half mesocoxal width (Fig. 31, 34); central macula on elytra yellowish-brown in both sexes; and elytra at least partially yellowish-brown posteriorly in both sexes. In the holotype female of *C. achiraense* (the only known specimen), the distance between the upper eye lobes is shorter, 0.3 times the distance between the outer margins of the eyes (Fig. 26), the distance between the lower eye lobes is 0.41 times the distance between the outer margins of the eyes, making the frons narrower (Fig. 29), the longitudinal carinae between the upper eye lobes are well marked (Fig. 29), the narrowest area of the mesoventral process is as wide as half the width of the mesocoxa (Fig. 27), central macula on elytra is yellowish white, and the elytra are entirely dark brown posteriorly. Additionally, the three specimens of *Compsibidion zonatum* examined have the first vertical row of the upper eye lobes with two ommatidia, while in there are three ommatidia in holotype female of *C. achiraensis*. However, this feature may eventually be variable. The new species is also similar to *C. amboroense* Santos-Silva, Galileo & Wappes, 2017 (see also photographs on Bezark (2025) and in Santos-Silva *et al.* (2017)), but differs especially by the elytra with more abundant, long and erect setae (very sparse in *C. amboroense*), yellowish-brown macula on center of elytra reaching the suture (not reaching in *C. amboroense*), inner area of the upper eye lobes narrower, with two ommatidia (wider, with three/four ommatidia in *C. amboroense*), and the widest area of the upper eye lobes with three ommatidia (five in *C. amboroense*).

Corrections in the species-group names

Note: In each species, the first species-group name is the one currently used; the second is the corrected one.

CERAMBYCIDAE Latreille, 1802

CERAMBYCINAE Latreille, 1802

TROPIDINI Martins & Galileo, 2007

NEOIBIDIONINA Monné, 2012

Compsibidion Thomson, 1864

Remarks. – *Compsibidion* is neuter gender because the suffix "-ίδιον" (-idion) is neuter and used to form neuter diminutive nouns. Therefore, two species-group names need to be corrected to agree with the gender of the genus:
– *Compsibidion achiraensis* Santos-Silva, Galileo & Wappes, 2017 – *C. achiraense* [the suffix "-ensis" is both masculine and feminine gender];
– *Compsibidion amboroensis* Santos-Silva, Galileo & Wappes, 2017 – *C. amboroense*.

New records

CERAMBYCIDAE Latreille, 1802

CERAMBYCINAE Latreille, 1802

TROPIDINI Martins & Galileo, 2007

COMPSINA Martins & Galileo, 2007

Paracompsa Martins, 2009

Paracompsa flavofasciata (Thomson, 1867)
(Fig. 37)

Ibidion (*Compsibidion*) *flavofasciatum* Thomson, 1867: 152.

Remarks. – This species was described based on syntypes from Colombia. Currently, it is known from Colombia and Venezuela (Monné 2024a; Tavakilian & Chevillotte 2024).

Material examined. – ECUADOR (**new country record**), *Imbabura*: Termas de Chachimbiro, 2600 m, **1** ♂, **1** ♀, 22 Jun 2023, J. Vlasak leg. (JVCO).

DISTENIIDAE Thomson, 1861

Arietocometes Santos-Silva & Tavakilian, 2009

Arietocometes violaceicollis (Villiers, 1958)
(Fig. 38)

Paracometes violaceicollis Villiers, 1958: 51.

Remarks. – This species was described and remains formally known only by the holotype and the two paratypes from Ecuador (Loja and Morona-Santiago) (Monné 2024b; Tavakilian & Chevillotte 2024).

Material examined. – ECUADOR, *Zamora-Chinchipe* (**new province record**): Zamora, 1000 m, **1** ♂, 7-8 Aug 2023, J. Vlasak leg. (JVCO).

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References

Bezark L.G., 2024. – Checklist of the Oxypeltidae, Vesperidae, Disteniidae and Cerambycidae (Coleoptera of the Western Hemisphere. 2024 Edition (updated through 31 December 2023). Available from: http://bezbycids.com/byciddb/wdefault.asp?w=n (Last accessed 08 January 2025).

Bezark L.G., 2025. – A photographic Catalog of the Cerambycidae of the World. New World Cerambycidae Catalogue. Available from: http://bezbycids.com/byciddb/wdefault.asp?w=n/ (Last accessed 08 January 2025).

Heffern D., Botero J.P. & Santos Silva A., 2020. – A new species of *Neocompsa* (Neoibidionini), and a new species and new combinations in Tillomorphini (Cerambycidae, Cerambycinae). *Zootaxa*, 4748(2): 334-350.

Lingafelter S.W., 2011. – New myrmecomorphous longhorned beetles from Haiti and the Dominican Republic with a key to Anaglyptini and Tillomorphini of Hispaniola (Coleoptera: Cerambycidae: Cerambycinae). *ZooKeys* 106: 55-75.

Martins U.R., 1975. – Longicórneos da coleção Hüdepohl III (Coleoptera, Cerambycidae). *Papéis Avulsos de Zoologia*, 29(2): 7-20.

Martins U.R. & Galileo M.H.M., 2013. – New species and records of Cerambycinae and Lamiinae (Coleoptera, Cerambycidae) from the Neotropical Region. *Zootaxa*, 3683(5): 571-580.

Monné M.A., 2024a. – Catalogue of the Cerambycidae (Coleoptera) of the Neotropical region. Part I. Subfamily Cerambycinae. Available from: http://cerambycids.com/default.asp?action=show_catalog/ (Last accessed 13 January 2025).

Fig. **37**-**38**. **37**. *Paracompsa flavofasciata* (Thomson, 1867), ♂, dorsal habitus. **38**. *Arietocometes violaceicollis* (Villiers, 1958), ♂, dorsal habitus.

Monné M.A., 2024b. – Catalogue of the Cerambycidae (Coleoptera) of the Neotropical region. Part II. Subfamily Lamiinae. Available from: http://cerambycids.com/default.asp?action=show_catalog/ (Last accessed 13 January 2025).

Monné M.A., 2024c. – Catalogue of the Cerambycidae (Coleoptera) of the Neotropical Region. Part III. Subfamilies Lepturinae, Necydalinae, Parandrinae, Prioninae, Spondylidinae, Anoplodermatinae and Families Oxypeltidae, Vesperidae and Disteniidae. Available from: http://cerambycids.com/default.asp?action=show_catalog/ (Last accessed 13 January 2025).

Monné M.A. & Nearns E.H., 2024. – Catalogue of the Cerambycidae (Coleoptera) of Canada and United States of America. Part III. Subfamily Cerambycidae. Available from: https://cerambycids.com/catalog/ (Last accessed 13 January 2025).

Roguet J.P., 2025. – Lamiinae of the World. Available: lamiinae.org/ (Last accessed 13 January 2025).

Santos-Silva A., Galileo M.H.M. & Wappes J.E., 2017. – Descriptions, notes and reassignments in Neoibidionini (Coleoptera:Cerambycidae: Cerambycinae) with a new genus, three new species and keys to species of *Brechmoidion* Martins, 1969, *Compsibidion* Thomson,1864 and *Rhysium* Pascoe, 1866. *Insecta Mundi*, 560: 1-21.

Tavakilian G.L. & Chevillotte H., 2024. – Titan: base de données internationales sur les Cerambycidae ou Longicornes. Available from: http://titan.gbif.fr/ (Last accessed 13 January 2025).

Thomson J., 1867. – Ibidionitarum species novae. *Physis Recueil d'Histoire Naturelle*, 1(3): 133-163.

Villiers A., 1958. – Révision du genre *Paracometes* Villiers (Col., Cerambycidae, Disteniinae). *Revue Française d'Entomologie*, 25: 43-56.

Vitali F., 2011. – Six new fossil Cerambycids included in Baltic and Saxon amber (Coleoptera Cerambycidae). *Entomapeiron* (P. S.), 4(1): 1-34.

Vitali F., 2017. – Two new fossil species of Tillomorphites Vitali (Coleoptera: Cerambycidae) and remarks on the morphological evolution, mimicry, biogeography and phylogeny of the tribe Tillomorphini. *Baltic Journal of Coleopterology*, 17(2): 147-140.

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Résumé

Vlasak J., Botero J. P. & Santos-Silva A., 2025. – Cerambycidae et Disteniidae (Coleoptera) d'Equateur : nouveaux genres, nouvelles espèces, nouvelles mentions et correction des noms de groupes d'espèces. *Faunitaxys*, 13(13) : 1 – 14.

Deux nouveaux genres et trois nouvelles espèces de Cerambycinae sont décrits d'Equateur : *Kerasellum afasciatum*, **gen**. **nov**., **sp**. **nov**. (Tillomorphini) ; *Afti citrinum*, **gen**. **nov**., **sp**. **nov**. (Trachyderini) ; et *Compsibidion zonatum* **sp**. **nov**. (Tropidini), ce dernier étant inclus dans une clé précédente. Deux noms de groupes d'espèces dans le genre *Compsibidion* Thomson, 1864 sont corrigés et deux nouvelles mentions géographiques sont signalées. L'inclusion actuelle d'*Eudereces elachys* Martins & Galileo, 2013 dans *Tillomorphites* Vitali, 2011, est remise en question dans les remarques sur *Kerasellum* **gen**. **nov**.

Mots-clés. – Coleoptera, Cerambycidae, Disteniidae, *Kerasellum*, *Afti*, *Compsibidion*, longicorne, description, taxonomie, nouveau genre, nouvelles espèces, Amérique du Sud.

Derniers articles publiés

Delahaye N., Marazzi G., Marazzi V. & Missori P., 2024. – Synonymy and description of a new species in *Xixuthrus* Thomson, 1864 from Indonesia (Coleoptera, Cerambycidae, Prioninae, Macrotomini, Xixuthrina). *Faunitaxys*, 12(42): 1 – 10. https://doi.org/10.57800/faunitaxys-12(42)

Vitali F., 2024. – Taxonomic notes about some Indo-Australian Lamiini (Coleoptera Cerambycidae). *Faunitaxys*, 12(43): 1 – 11. https://zoobank.org/4D315C8D-2EB6-4F66-819D-B606FF4ACDA3

Fei-Yi Xin, Yi-Feng Zhang & Zhi-Hao Qi, 2024. – *Falcicornis zhongi* Xin & Qi, **sp**. **nov**., a new stag beetle species from Vietnam (Coleoptera: Lucanidae: Lucaninae). *Faunitaxys*, 12(44): 1 – 6. https://doi.org/10.57800/faunitaxys-12(44)

Lanuza-Garay A., Santos-Murgas A. & Cambra R.A., 2024. – Checklist of True Weevils (Coleoptera: Curculionidae) from Rancho Frio, Darien National Park, Panama. *Faunitaxys*, 12(45): 1 – 7. https://doi.org/10.57800/faunitaxys-12(45)

Lourenço W. R. & Velten J., 2024. – New insights on the diversity of the family Chaerilobuthidae Lourenço & Beigel, 2011 with the description of one new genus and species (Scorpiones). *Faunitaxys*, 12(46): 1 – 6. https://doi.org/10.57800/faunitaxys-12(46)

Platia G. & Konvička O., 2024. – Description of three new species of click beetles (Coleoptera: Elateridae) with an updated catalogue of Elateridae from the Sultanate of Oman. *Faunitaxys*, 12(47): 1 – 11. https://doi.org/10.57800/faunitaxys-12(47)

Háva J., 2024. – A new *Mycetophagus* (subgenus *Ilendus* Casey, 1900) species from Iran (Coleoptera: Mycetophagidae). *Faunitaxys*, 12(48) : 1 – 3. https://doi.org/10.57800/faunitaxys-12(48)

Lourenço W. R. & Velten J., 2024. – Possible estimation of litter size in palaeoburmesebuthid scorpions from Early Cretaceous Burmite (Chelicerata: Scorpiones). *Faunitaxys*, 12(49): 1 – 4. https://doi.org/10.57800/faunitaxys-12(49)

Vives E., 2024. – Lepturinae nuevos o interesantes del sudeste asiático (Coleoptera, Cerambycidae). Notes on Lepturinae (22). *Faunitaxys*, 12(50) : 1 – 9. https://doi.org/10.57800/faunitaxys-12(50)

Lapèze J., 2024. – Synthèse des connaissances sur les membracides et les aetalions de Guyane Française (Hemiptera : Auchenorrhyncha : Membracidae, Aetalionidae). Partie I : historique et liste des espèces recensées. *Faunitaxys*, 12(51) : 1 – 35. https://doi.org/10.57800/faunitaxys-12(51)

Gereys B., 2024. – Examen critique des synonymies des Vespidae sociaux de France métropolitaine (Hymenoptera : Vespidae) – Partie 2 (Vespinae). *Faunitaxys*, 12(52) : 1 – 39. https://doi.org/10.57800/faunitaxys-12(52)

Santos-Silva A., Bezark L. G. & Tejada Hurtado G., 2024. – New records of South American Cerambycidae (Coleoptera), with notes on the authorship of *Homogenes* and corrections in the species-group names of some species. *Faunitaxys*, 12(53): 1 – 8. https://doi.org/10.57800/faunitaxys-12(53)

Keith D., 2024. – Redécouverte d'*Anomala papuna* Arrow, 1917, espèce orientale méconnue (Coleoptera: Scarabaeoidea, Scarabaeidae, Rutelinae). *Faunitaxys*, 12(54): 1 – 3 https://doi.org/10.57800/faunitaxys-12(54)

Vincent B., 2024. – Description de trois nouvelles espèces d'altitude appartenant au genre *Amastus* Walker, 1865 (Lepidoptera: Erebidae, Arctiinae). *Faunitaxys*, 12(55) : 1 – 7. https://doi.org/10.57800/faunitaxys-12(55)

Lourenço W. R. & Velten J., 2024. – The remarkable diversity of the genus *Chaerilobuthus* Lourenço & Beigel, 2011 with the description of one more new species (Scorpiones: Chaerilobuthidae). *Faunitaxys*, 12(56): 1 – 5. https://doi.org/10.57800/faunitaxys-12(56)

Hui Wen, Xin-Yuan Zhang, Shu-Tong Zhang & Hao Xu, 2024. – Distribution of the flower beetle *Pseudodiceros nigrocyaneus* (Bourgoin, 1917) (Coleoptera: Scarabaeidae: Cetoniinae) in Asia with biological notes. *Faunitaxys*, 12(57): 1 – 15. https://doi.org/10.57800/faunitaxys-12(57)

Cumming R. T., Engel M. S., Lian Z. & Ulitzka M. R., 2024. – Small, intricate, and beautiful; a new species of lophioneurid from the Cretaceous (Insecta: Thripida: †Lophioneurida). *Faunitaxys*, 12(58): 1 – 10. https://doi.org/10.57800/faunitaxys-12(58)

Ythier E. & Lourenço W. R., 2024. – A new species of *Hadrurochactas* Pocock, 1893 (Scorpiones: Chactidae) from the Mitaraka Massif in French Guiana. *Faunitaxys*, 12(59): 1 – 9. https://doi.org/10.57800/faunitaxys-12(59)

Háva J., 2024. – *Mycetophagus* (s. str.) *lenkae* **sp**. **nov**. from India: Arunachal Pradesh (Coleoptera: Mycetophagidae), along with new records for *Mycetophagus* (*Mycetophagoides*) *alni* Champion in two states in India. *Faunitaxys*, 12(60) : 1 – 3. https://doi.org/10.57800/faunitaxys-12(60)

Herbin D., Giusti A. & Mielke C. G. C., 2024. – Taxonomic revision of the genera *Tamphana* Schaus and *Tuyvae* Orlandin, Piovesan & Carneiro with description of eleven new species (Lepidoptera, Apatelodidae). *Faunitaxys*, 12(61): 1 – 31. https://doi.org/10.57800/faunitaxys-12(61)

Niisato T. & Tichý T., 2024. – A new species of the genus *Microdebilissa* Pic, 1925 from West Kalimantan (Coleoptera, Cerambycidae, Cerambycinae, Stenopterini). *Faunitaxys*, 12(62): 1 – 4. https://doi.org/10.57800/faunitaxys-12(62)

Moret P., 2024. – Description of new Platynini from the montane cloud forest of Ecuador, with a redefinition of the genera *Glyptolenus* Bates and *Glyptolenoides* Perrault (Coleoptera, Carabidae). *Faunitaxys*, 12(63): 1 – 23. https://doi.org/10.57800/faunitaxys-12(63)

Cerda J.-A., 2024. – Corrigenda de *Butlerius* et *Guerinius* (Lepidoptera, Noctuoidea, Erebidae, Arctiinae, Arctiini, Ctenuchina). Quinzième note. *Faunitaxys*, 12(64) : 1. https://doi.org/10.57800/faunitaxys-12(64)

Vives E., 2024. – Un nuevo género de Strogylurini de las islas Filipinas. New or interesting Cerambycidae from Philippines (Col.: Cerambycidae; Cerambycinae) (Pars 31). *Faunitaxys*, 12(65) : 1 – 4. https://doi.org/10.57800/faunitaxys-12(65)

Vlasak J., Santos-Silva A. & Bezark L. G., 2024. – South American Acanthocinini (Coleoptera, Cerambycidae, Lamiinae): description of a new species; transfer of a species from *Urgleptes* to *Lepturges* (*Lepturges*); designation of a lectotype; and corrections. *Faunitaxys*, 12(66) : 1 – 11. https://doi.org/10.57800/faunitaxys-12(66)

Bollino M. & Racheli T., 2025. – A new taxon of the *Pterourus menatius* complex from West Ecuador (Lepidoptera: Papilionidae). *Faunitaxys*, 13(01): 1 – 6. https://doi.org/10.57800/faunitaxys-13(01)

Lourenço W. R. & Velten J., 2025. – One more new species for the genus *Cretaceoushormiops* Lourenço, 2018 from Cretaceous Burmite (Scorpiones: Protoischnuridae). *Faunitaxys*, 13(02): 1 – 6. https://doi.org/10.57800/faunitaxys-13(02)

Lourenço W. R. & Velten J., 2025. – A new species for the genus *Archaeoananteroides* Lourenço, 2016, fossil scorpion from Burmite (Scorpiones: Buthoidea: Buthidae). *Faunitaxys*, 13(03): 1 – 6. https://doi.org/10.57800/faunitaxys-13(03)

Lourenço W. R. & Ythier E., 2025. – A new cave species of *Isometrus* Ehrenberg 1828 from Myanmar (Scorpiones: Buthidae). *Faunitaxys*, 13(04): 1 – 6. https://doi.org/10.57800/faunitaxys-13(04)

Platia G. & Pedroni G., 2025. – Five new species and new records of click beetles from Saudi Arabia with an updated catalogue (Coleoptera, Elateridae). *Faunitaxys*, 13(05): 1 – 12. https://doi.org/10.57800/faunitaxys-13(05)

Háva J. & Hodeček J., 2025. – Two additional new species of *Cryptorhopalum* (Coleoptera: Dermestidae: Megatominae) from the Dominican Republic. *Faunitaxys*, 13(06): 1 – 5. https://doi.org/10.57800/faunitaxys-13(06)

Vitali F., 2025. – The genus *Epicasta* Thomson, 1864 and the limits of the morphological systematics in the study of Lamiinae (Coleoptera Cerambycidae). *Faunitaxys*, 13(07): 1 – 5. https://doi.org/10.57800/faunitaxys-13(07)

Ythier E., Sadine S. E., Alioua Y. & Lourenço W. R., 2025. – A new species of *Androctonus* Ehrenberg, 1828 from the Tassili n'Ajjer, Algeria (Scorpiones: Buthidae). *Faunitaxys*, 13(08): 1 – 8. https://doi.org/10.57800/faunitaxys-13(08)

Strümpher W. P. & Stals R., 2025. – A world list of known cave- and bat guano-associated Trogidae (Coleoptera: Scarabaeoidea), and further range extension of *Omorgus lindemannae* (Petrovitz). *Faunitaxys*, 13(09): 1 – 11. https://doi.org/10.57800/faunitaxys-13(09)

Lourenço W. R. & Velten J., 2025. – Description of a second species for the genus *Serratochaerilobuthus* Lourenço, 2024 (Scorpiones: Chaerilobuthidae). *Faunitaxys*, 13(10): 1 – 6. https://doi.org/10.57800/faunitaxys-13(10)

Jarzabek-Müller A., 2025. – A new *Idolus* Desbrochers des Loges, 1875 for the Caucasus region (Coleoptera, Elateridae, Pomachiliini). *Faunitaxys*, 13(11): 1 – 6. https://doi.org/10.57800/faunitaxys-13(11)

Laguerre M., 2025. – New species of Arctiinae east of the Canal Zone, Panama (Lepidoptera, Erebidae, Arctiinae, Arctiini). *Faunitaxys*, 13(12): 1 – 17. https://doi.org/10.57800/faunitaxys-13(12)

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Cerambycidae et Disteniidae (Coleoptera) d'Equateur : nouveaux genres, nouvelles espèces, nouvelles mentions et correction des noms de groupes d'espèces.

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